RayBio® G-Series Rat Protein Tyrosine Phosphorylation Antibody Array 3

For Simultaneously Detecting the Relative Level of Tyrosine Phosphorylation of Rat Protein

User Manual

(Revised Mar. 20th, 2024)

Cat#: AAR-PTYR-G3-4 (4 Sample Kit)
Cat#: AAR-PTYR-G3-8 (8 Sample Kit)

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Empowering your proteomics

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Tel: (770) 729-2992 **Fax: (888) 547-0580**
Web: www.raybiotech.com **Email: info@raybiotech.com**

RayBiotech Life, Inc.

RayBio® G-Series Rat Protein Tyrosine Phosphorylation Antibody Array 3 Protocol

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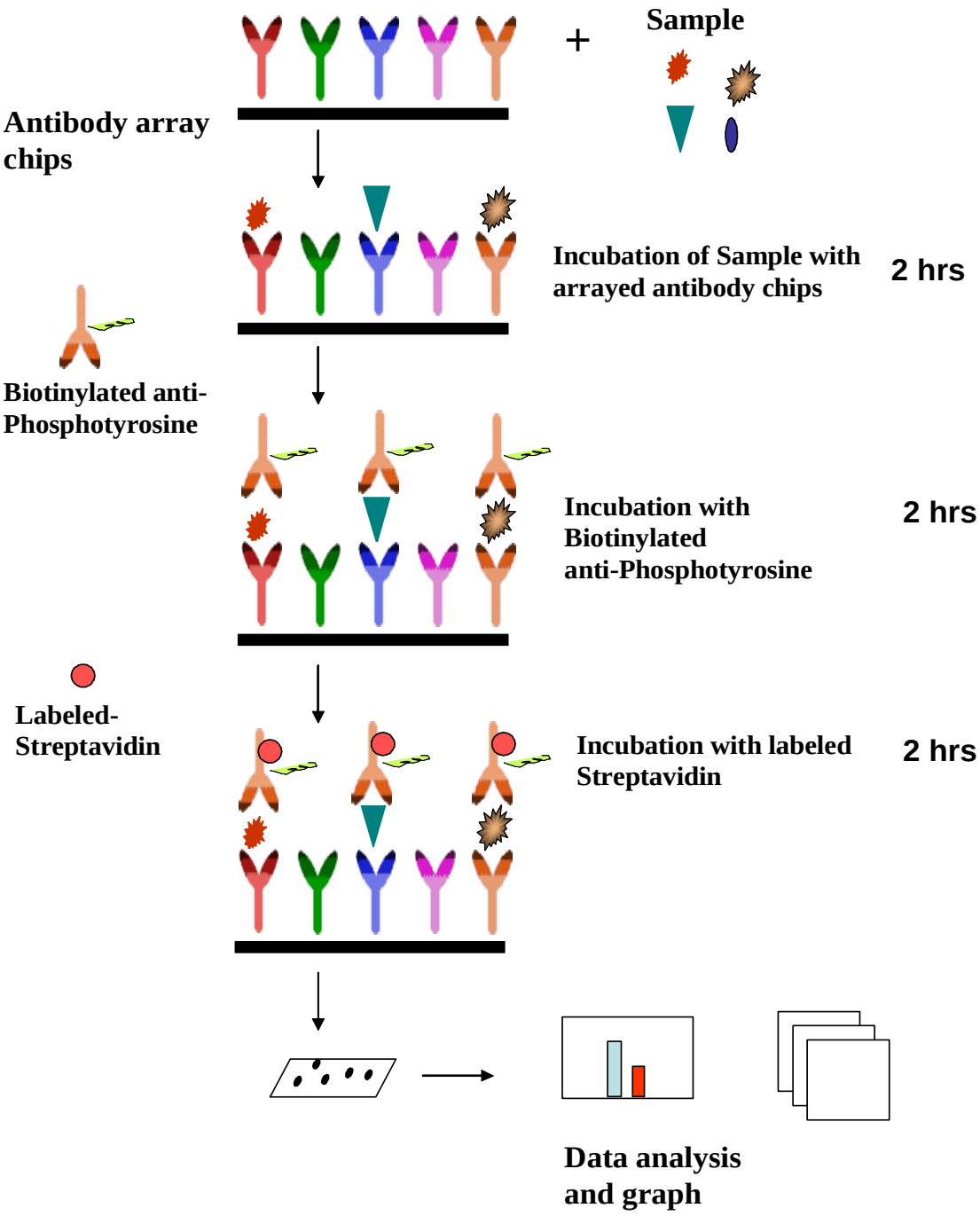
I. Introduction

Protein phosphorylation plays an unusually prominent role in cell signaling, development and growth. The RayBio® G-Series Rat Protein Tyrosine Phosphorylation Antibody Array 3 is a very rapid, convenient, and sensitive assay that can simultaneously detect multiple protein phosphorylations and be used to monitor the activation or function of important biological pathways.

RayBiotech is committed to develop a series of phosphorylation antibody arrays. RayBio® Rat Protein Tyrosine Phosphorylation Antibody Array 3 is specifically designed for simultaneous identification of the relative levels of phosphorylation of 500 different Rat Proteins in cell lysate. By monitoring the changes in protein tyrosine phosphorylation in your experimental model system, you can verify pathway activation in your cell lines without spending excess time and effort performing an analysis of immunoprecipitation and/or Western Blot.

To use the RayBio® G-Series Rat Protein Tyrosine Phosphorylation Antibody Array 3, treated or untreated cell lysate is added into antibody array glass slide wells. The antibody array slide wells are washed, and biotinylated anti-phosphotyrosine antibodies are then used to detect the phosphorylated tyrosines on target proteins. After incubation with a fluorescent dye-conjugated streptavidin (Cy3 equivalent), the slides can then be imaged using a laser scanner, such as the Axon GenePix, using the Cy3 channel.

Here's how it works



II. Materials Provided

Store kit at ≤ -20 °C immediately upon arrival. Kit must use within the 6 months expiration date.

ITEM	COMPONENT	AAR-PTYR-G3-4	AAR-PTYR-G3-8	STORAGE TEMPERATURE AFTER THAWING**
1	RayBio® Glass Slide*	1	2	≤-20°C
2	Blocking Buffer	1 bottle (8ml/ea)	2 bottles (8ml/ea)	
3	Biotinylated Anti- PhosphoTyrosine Antibody	1 vial	2 vials	2-8°C
4	Cy3 equivalent-Conjugated Streptavidin	1 vial	2 vials	2-8 °C
5	20X Wash Buffer I Concentrate	1 bottle (30 ml)		2-8 °C
6	20X Wash Buffer II Concentrate	1 bottle (30 ml)		
7	Wash Buffer III	1 bottle (20 ml)		
8	2X Cell Lysis Buffer Concentrate	1 bottle (10ml)		2-8 ℃
9	Protease Inhibitor Cocktail	1 vial		≤-20°C
10	Phosphatase Inhibitor Cocktail II	1 vial		
Other Kit Components: Adhesive film				

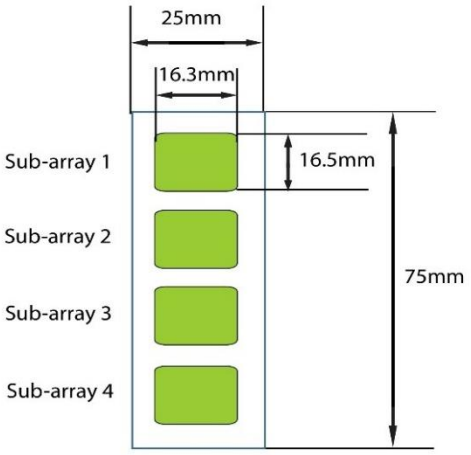
*Each slide contains 4 identical subarrays

**For up to 3 months (unless stated otherwise) or until expiration date

III. Additional Materials Required

- Shaker
- Laser scanner for fluorescence detection
- Aluminum foil
- Distilled water
- Plastic box
- 50 ml Centrifuge tube
- Isopropanol (2-propanol)

Layout of Array Glass Slide



4 printed sub-arrays per glass chip

IV. Reagent Preparation

1. **Protease Inhibitor Cocktail:** Briefly spin down the Protease Inhibitor Cocktail vial before use. Add 60 µl of 1X Cell Lysis Buffer to the vial to prepare a 100X Protease Inhibitor Cocktail Concentrate.

2. **Phosphatase Inhibitor Cocktail Set II:** Briefly spin down the Phosphatase Inhibitor Cocktail Set II vial before use. Add 180 µl of 1X Cell Lysis Buffer to the vial to prepare a 25X Phosphatase Inhibitor Cocktail Set II Concentrate**. Dissolve the powder thoroughly by gentle mixing.**

3. **2X Cell Lysis Buffer:** The 2X Cell Lysis Buffer should be diluted 2-fold with deionized or distilled water to prepare a 1X Cell Lysis Buffer solution. Then, add 20 µl of the Protease Inhibitor Cocktail Concentrate and 80 µl of the Phosphatase Inhibitor Cocktail Set II Concentrate into 1.9 ml of the 1X Cell Lysis Buffer to prepare a 1X Cell Lysis Buffer with Protease and Phosphatase Inhibitor Cocktail solution. Mix well before use.

4. **20X Wash Buffer I or II:** If the 20X Wash Buffer Concentrate contains visible crystals, warm to room temperature and mix gently until dissolved. Dilute 25 ml of the 20X Wash Buffer Concentrate into deionized or distilled water to yield 500 ml of 1X Wash Buffer.

5. **Biotinylated anti-Phosphotyrosine:** Briefly spin down the Detection Antibody vial before use. Add 90 µl of Blocking Buffer to the vial to prepare a Biotinylated Anti-phosphotyrosine Concentrate. Pipette up and down to mix gently (the Concentrate can be stored at 4 ^{0}C for 5 days). Add 90 µl of Detection Antibody Concentrate to a tube with 1710 µl of Blocking Buffer to prepare a 1X Biotinylated Anti-phosphotyrosine solution. Mix gently.

6. **Fluorescent dye-Conjugated Streptavidin (Cy3 equivalent):** Briefly spin down the Fluorescent dye-Conjugated Streptavidin vial before use. Add 180 µl of Blocking Buffer to the vial to prepare a Streptavidin

Concentrate. Pipette up and down to mix gently. Transfer all Streptavidin Concentrate to a tube with 1.7 ml of Blocking Buffer to prepare a 1X Fluorescent dye-Conjugated Streptavidin solution. Mix gently.

V. Overview and General Considerations

A. Preparation of Samples

Cells can be prepared using the following convention.

For attached cells, remove the supernatant from the cell culture, and wash the cells twice with cold 1X PBS (for cells in suspension, pellet the cells by spinning down at 1500 rpm for 10 min). Make sure to remove any remaining PBS. Then, solubilize the cells at $2x10^7$ cells/ml in the 1X Cell Lysis Buffer with Protease and Phosphatase Inhibitor Cocktail solution. Pipette up and down to resuspend the cells, and rock the lysates gently at 2–8 °C for 30 min. Transfer the lysates to microcentrifuge tubes and centrifuge at 14,000 x g for 5 min.

It is recommended that sample protein concentrations be determined using a total protein assay. For incubation with the Phosphorylation Antibody Array G-series 1, use cell lysates at a concentration of 50–1000 µg/ml (as a starting point, we recommend using 400 µg/ml of cell lysate diluted at least 5-fold with the Blocking Buffer).

Lysates should be used immediately or aliquoted and stored at –80 °C. Thawed lysates should be kept on ice prior to use.

If you experience high background, you may further dilute your sample.

B. Handling glass slides

- The microarray slides are very sensitive. Do not touch the array surface with tips, forceps or hands. Hold the slides by the edges only.

- Handle all buffers and slides with latex free gloves.
- Avoid breaking the glass slide.
- Maintain a clean environment.

C. Incubation

- Completely cover the array area with sample or buffer during incubation, and cover the incubation chamber with the adhesive film or plastic sheet protector to avoid drying.
- Avoid foaming during incubation steps.
- Perform all incubation and wash steps under gentle rotation.
- Cover the incubation chamber with the adhesive film during incubation, particularly when the incubation is more than 2 hours.
- Avoid cross-contamination from overflowing solution to neighboring wells.
- Several incubation steps such as step 2 (sample incubation), step 6 (Biotin-conjugated Anti-phosphotyrosine incubation) or step 9 (Fluorescent dye-Conjugated Streptavidin incubation) may be done at 4 °C overnight. Please make sure to cover the incubation chamber tightly to prevent evaporation.
- Avoid exposing the array slide to light from step 9 in page 10 on.

VI. Protocol

A. Dry the Glass Slide

Open the box containing the Glass Slide with Frame and take it out. Then let it air dry for 1 hour in a clean environment before use.

Note: *Protect the slide from dust or other contaminants.*

B. Blocking and Incubation

1. Add 400 µl of 1X Blocking Buffer to each well and incubate at room temperature with gentle shaking for 30 min to block the slides. Make sure no bubbles are in the wells.

2. Decant the Blocking Buffer from each well (make sure to remove all of the buffer). Add 400 µl of each sample into appropriate wells. Incubate the arrays with sample at room temperature with gentle shaking for 2 hours or at 4 °C overnight.

   Note: We recommend using 400 µl of cell lysate at a concentration of 50–1000 ug/ml (as a starting point, we recommend using 400 µg/ml cell lysate). ***Dilute the lysate at least 5-fold with the Blocking Buffer. Make sure there are no bubbles in the wells.***

   Note: The amount of sample used depends on the abundance of target proteins. More sample can be used if signals are too weak. If signals are too strong, the sample can be diluted further. The optimal sample dilution must be determined empirically by the researcher.

   Note: Incubation may be done at 4 °C overnight.

3. Decant the samples from each well, and wash 3 times, 5 min per wash, with 800 µl of 1X Wash Buffer I at room temperature with gentle shaking.

   Note: Avoid the solution overflowing into neighboring wells.

4. Put the Glass Slide with Frame into a box with Wash Buffer I (cover the whole glass slide and frame with Wash Buffer I), and wash at room temperature with gentle shaking for 20 min.

5. Decant the Wash Buffer I from each well. Put the Glass Slide with Frame into a box with Wash Buffer II (cover the whole glass slide and frame with Wash Buffer II), and wash 2 times, 5 min per wash, at room temperature with gentle shaking.

6. Remove all of Wash Buffer II from each well. Add 400 µl of the 1X Biotin-conjugated Anti-phosphotyrosine solution to each corresponding well. Incubate at room temperature with gentle shaking for 2 hours.

7. Decant the antibody solution and wash as directed in step 4 three times (wash 3 times, 20 min per wash).

8. Wash as directed in step 5.

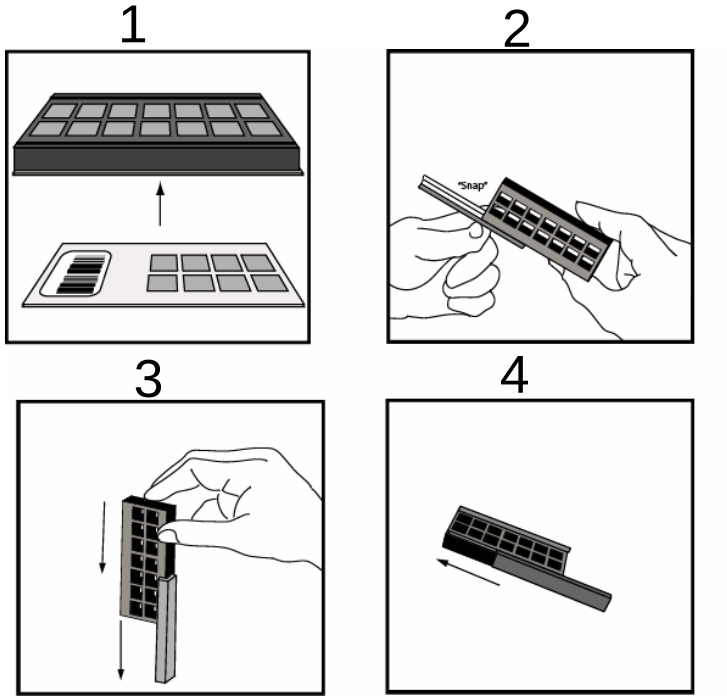
9. Remove all of Wash Buffer II from each well. Add 400 µl of the 1X Fluorescent dye-Conjugated Streptavidin solution to each subarray. Cover the incubation chamber with the Adhesive film. Cover the plate with aluminum foil to avoid exposure to light or incubate in a dark room.

   Note: Avoid exposing the array slide to light from this step forward.

10. Incubate at room temperature with gentle shaking for 2 hours in the dark.

Note: Incubation may be done at 4 °C overnight.

11. Decant the Fluorescent dye-Conjugated Streptavidin solution and disassemble the Glass Slide and Frame by removing the incubation frame and chamber from the slide as illustrated below.



Note: You may assemble and disassemble the glass slide into an incubation chamber and glass slide using the following steps.

1. To assemble, apply the incubation chamber to the slide with the printed side facing upward as illustrated in (1) above.

2. Gently snap one edge of a snap-on side as shown in (2).

3. Adjust the position of the snap-on by gently pressing the edge of the snap-on side against a lab bench and pushing down as shown in (3).

4. Repeat steps 2 – 3 with a second snap-on as shown in (4).

12. Gently put the glass slide into a 50 ml centrifuge tube or a plastic box with 40 ml of 1X Wash Buffer I as illustrated below. Gently roll or shake the tube for 5 min. Remove the Wash Buffer I. Repeat 2 more times for a total of 3 washes.

13. Wash the glass slide with 40 ml of Wash Buffer II for 5 min. Repeat one more time for a total of 2 washes.

14. Finally, wash the glass slide with 40 ml of deionized or distilled water.

C. Fluorescence Detection

1. To dry the glass slide, do one of the following:
    a. Put the glass slide into a 50 ml centrifuge tube and centrifuge at 1,000 rpm for 3 min
        or
    b. Apply a compressed N_2 stream, or let glass slide air dry completely under clean air conditions (protected from light)

    Make sure the slides are absolutely dry before scanning.

2. Image the slides using a laser scanner, such as the Axon GenePix, using the Cy3 channel.

    Note: We recommend scanning the slides immediately after completing the experiment. Slides can also be stored at –20 °C in the dark for

several days. If you do not have a laser scanner, we can scan and extract the data for free for you.

Note: Put the glass slide into a tube with 40 ml of 30% Wash Buffer III in isopropanol (add 15 ml of Wash Buffer III to a tube with 35 ml of isopropanol and mix well) and incubate for 10 min at room temperature if the background is not even or too high (cover the tube with aluminum foil to avoid exposure to light or incubate in a dark room). Dry the slide completely and re-scan the slide.

VII. Interpretation of Results

The following figure shows the RayBio® G-Series Rat Protein Tyrosine Phosphorylation Antibody Array 3 probed with different cell lysates. The images were captured using a laser scanner. A biotinylated protein produces positive control signals, which can be used to identify the orientation of the slide and to normalize the results for comparison of different wells.

The antibody affinity to its target varies significantly between different antibodies. The fluorescence intensity detected on the array with each antibody depends on this affinity; therefore, the signal intensity comparison can only be performed within the same antibody/antigen system and not between different antibodies on the same slide. Certain proteins containing phosphorylated tyrosine may not be recognized by biotinylated anti-phosphotyrosine because of steric hindrance of the recognition site.

RayBio® G-Series Rat Protein Tyrosine Phosphorylation Antibody Array 3 Array Map

	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30
1	POS1	POS1	POS2	POS2	POS3	POS3	Neg	Neg	1	1	2	2	3	3	4	4	5	5	6	6	7	7	8	8	9	9	10	10	11	11
2	12	12	13	13	14	14	15	15	16	16	17	17	18	18	19	19	20	20	21	21	22	22	23	23	24	24	25	25	26	26
3	27	27	28	28	29	29	30	30	31	31	32	32	33	33	34	34	35	35	36	36	37	37	38	38	39	39	40	40	41	41
4	42	42	43	43	44	44	45	45	46	46	47	47	48	48	49	49	50	50	51	51	52	52	53	53	54	54	55	55	56	56
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7	87	87	88	88	89	89	90	90	91	91	92	92	93	93	94	94	95	95	96	96	97	97	98	98	99	99	100	100	101	101
8	102	102	103	103	104	104	105	105	106	106	107	107	108	108	109	109	110	110	111	111	112	112	113	113	114	114	115	115	116	116
9	117	117	118	118	119	119	120	120	121	121	122	122	123	123	124	124	125	125	126	126	127	127	128	128	129	129	130	130	131	131
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11	147	147	148	148	149	149	150	150	151	151	152	152	153	153	154	154	155	155	156	156	157	157	158	158	159	159	160	160	161	161
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20	POS1	POS1	POS2	POS2	POS3	POS3	Neg	Neg	282	282	283	283	284	284	285	285	286	286	287	287	288	288	289	289	290	290	291	291	292	292
21	293	293	294	294	295	295	296	296	297	297	298	298	299	299	300	300	301	301	302	302	303	303	304	304	305	305	306	306	307	307
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33	473	473	474	474	475	475	476	476	477	477	478	478	479	479	480	480	481	481	482	482	483	483	484	484	485	485	486	486	487	487
34	488	488	489	489	490	490	491	491	492	492	493	493	494	494	495	495	496	496	497	497	498	498	499	499	500	500	Neg	Neg	Neg	Neg
35	Neg	Neg	Neg	Neg	Neg	Neg	Neg	Neg	Neg	Neg	Neg	Neg	Neg	Neg	Neg	Neg	Neg	Neg	Neg	Neg	Neg	Neg	Neg	Neg	POS3	POS3	POS2	POS2	POS1	POS1

RayBio® Rat Protein Tyrosine Phosphorylation Antibody Array G3 Target List

Number	Name	Number	Name	Number	Name	Number	Name	Number	Name	Number	Name	Number	Name
1	A2M	73	CHGB	145	FGFR5	217	IL-1 R4	289	Neurturin	361	Quiescin Q6	433	TCP1
2	AARE	74	Chk1	146	FGG	218	IL-1 R6	290	Nidogen-1	362	RAGE	434	TDIF2
3	ABCF1	75	Chymase	147	FH	219	IL-11	291	Nidogen-2	363	Ras	435	TECK
4	ACAT1	76	CINC-2	148	Fibronectin	220	IL-12 p40	292	NIT2	364	RELM beta	436	Tenascin X
5	Activin A	77	CINC-3	149	Ficolin-2	221	IL-12 RB1	293	NNT	365	Resistin	437	TFF1
6	ADAMTS10	78	Cingulin	150	FLG2	222	IL-13	294	NOV	366	REV3L	438	TFF2
7	ADAMTS15	79	CIP29	151	FOXN3	223	IL-15	295	NPB	367	Rheb	439	TGF-beta 1
8	ADAMTSL2	80	Claudin-3	152	Fractalkine	224	IL-16	296	NPTXR	368	RNASE6	440	TGF-beta 2
9	Aggrecan	81	Claudin-4	153	Frizzled-1	225	IL-17A	297	NR3C3	369	ROBO4	441	TGF-beta 3
10	AHCY	82	CNPY2	154	Frizzled-4	226	IL-17C	298	Nrf1	370	ROR1	442	TGF-beta R1
11	AHSG	83	CNTFR	155	Frizzled-5	227	IL-17D	299	OCT3/4	371	RP1	443	TGF-beta R2
12	Akt2	84	COL19A1	156	Frizzled-6	228	IL-19	300	Orexin A	372	RPL12	444	TIMP-1
13	Albumin	85	COTL1	157	Frizzled-7	229	IL-2 R beta	301	OSCAR	373	RPL23A	445	TIMP-2
14	AMPKa1	86	CPE	158	FSTL1	230	IL-24	302	OSM	374	RPLP0	446	Titin
15	ANGPTL2	87	CRADD	159	Galanin	231	IL-27	303	Osteoactivin	375	RPS13	447	TK1
16	ANGPTL3	88	CREB	160	GASP-1	232	IL-28B	304	Osteoadherin	376	RPS14	448	TLR1
17	ANKRD9	89	CRF21	161	GASP-2	233	IL-3	305	Osteoprotegerin	377	RPS15A	449	TLR3
18	ANXA6	90	CRHBP	162	G-CSF R	234	IL-3 R beta	306	p130Cas	378	RPS23	450	TLR4
19	APBA2	91	CrkRS	163	GDF-15	235	IL-5	307	p21	379	RPS3A	451	TMEFF1
20	ApoA1	92	CRTAC1	164	GDF-5	236	IMP2	308	P4HB	380	RPS5	452	TMEFF2
21	ApoA2	93	CRTAM	165	GFRA4	237	INSL3	309	Pappalysin-1	381	RPS8	453	TMEM223
22	ApoB	94	CRTH-2	166	GHR	238	Inuslin	310	PCAP	382	RPS9	454	TOMM70A
23	ApoE	95	Cryptic	167	GKN1	239	I-TAC	311	PCPE-1	383	RREB1	455	TPIS
24	ARHGAP1	96	CSE1L	168	GLI-2	240	Jak2	312	PD-1	384	RSF1	456	TPP1
25	ATG5	97	CSK	169	GLIPR2	241	Kallikrein 10	313	PD-ECGF	385	RUSC1	457	TRADD
26	ATPG	98	CTNND1	170	Glut1	242	Kallikrein 11	314	PDGF-AA	386	S100A10	458	TRAIL R2
27	B3GAT1	99	CXCR2	171	Glut2	243	Kallikrein 5	315	PDGF-C	387	S100A11	459	TRAM
28	B4GalT1	100	CXCR4	172	Glut4	244	Kallikrein 6	316	PDGF-D	388	S100A9	460	TRIM14
29	B7-1	101	CXCR7	173	Glut5	245	KIF5B	317	PDGFRB	389	S-100b	461	Tropomyosin 3
30	B7-H2	102	Cyclin D1	174	GM2A	246	LAMA5	318	PDLIM5	390	SBP-1	462	TRRAP
31	BAFF R	103	Cyclophilin F	175	GM-CSF	247	LAMP	319	PDZD2	391	SCF	463	Trypsinogen-2
32	Bax	104	Cystatin A	176	GP2	248	LASP1	320	PENK	392	SCF R	464	TSLP
33	BDNF	105	Cystatin B	177	gp340	249	LBP	321	Pentraxin-3	393	SDF4	465	TSP-1
34	beta-NGF	106	Cystatin D	178	GPD1	250	Lefty-1	322	Perilipin-3	394	Septin-7	466	TSP-2
35	BLAME	107	Cystatin E	179	GPR-39	251	Lefty-A	323	Peroxiredoxin-3	395	SERBP1	467	TSP-4
36	BLMH	108	Cystatin S	180	Granzyme A	252	LHPP	324	PF4	396	Serpin A3	468	TTF1
37	BMP-1	109	DAK	181	Granzyme M	253	LIX	325	PFAS	397	Serpin A5	469	TUBA6
38	BMP-15	110	DCI	182	GRHPR	254	LPS	326	PFDN6	398	Serpin B5	470	TWF2
39	BMP-9	111	DCXR	183	GRP	255	LRG1	327	PHGDH	399	Serpin C1	471	TXNDC15
40	BNIP2	112	DLL4	184	GSK-3 beta	256	LRP-6	328	Piccolo	400	SET	472	TXNDC5
41	BOLA2	113	DMGDH	185	GSN	257	L-Selectin	329	PIK3R2	401	sFRP-4	473	TYRO10
42	BTC	114	DSCAM	186	GSR	258	LUZP1	330	PINCH1	402	SH3BGRL3	474	UBC9
43	BTF3	115	DSG1	187	GSTM1	259	Lymphotactin	331	PIP4K2A	403	SHBG	475	Ubiquitin
44	C1q	116	EDA-A2	188	GSTO1	260	MAdCAM-1	332	PLA2G1B	404	SHOX	476	Ubiquitin+1
45	C1s	117	EDAR	189	GULP1	261	MAN1	333	PLD4	405	Siglec-1	477	UNC45A
46	C3a	118	eEF2	190	HAI-1	262	Mcl-1	334	Plexin B2	406	SLC38A10	478	UNC5H4
47	C5a	119	EG-VEGF	191	Haptoglobin	263	MCP-1	335	PIGF-2	407	SLITRK1	479	uPA
48	CA1	120	eIF4E	192	HB-EGF	264	MCP-5	336	PLS3	408	SLPI	480	UROC1
49	CA2	121	EMAP-II	193	HEG1	265	MDC	337	PNP	409	SLURP1	481	USP2
50	CA3	122	Endothelin	194	Hepassocin	266	MEP1A	338	POMC	410	Smad 1	482	Uteroglobin
51	Calbindin D	123	Eotaxin-2	195	HEXB	267	Mesothelin	339	PON1	411	Smad 4	483	VAP-1
52	Cardiotrophin-1	124	EphA1	196	HGFA	268	MICB	340	PP	412	Smad 5	484	VAP-A
53	Cathepsin A	125	EphA2	197	Histone H2AY	269	MIP-3 alpha	341	PPP1CC	413	Smad 8	485	VARS
54	CCL28	126	EPHX2	198	hnRNPL	270	MIS RII	342	PRAT4B	414	Somatostatin	486	VDAC1
55	CCR3	127	Epiregulin	199	Hoxb3	271	Mitofusin 2	343	PRELP	415	SOX5	487	VEGF
56	CCR4	128	ERRa	200	HOXD11	272	MKK3	344	Prolactin R	416	SPARC	488	VEGF-B
57	CCT3	129	E-Selectin	201	HSP10	273	MKK4	345	ProSAAS	417	SPINK7	489	VEGF-C
58	CD133	130	EVC2	202	HSP47	274	MMP-10	346	Prostasin	418	SPTBN5	490	VEGFR3
59	CD23	131	Factor IX	203	HTRA1	275	MMP-13	347	Protein Z	419	SSTR2	491	VILIP3
60	CD24	132	Factor V	204	HVEM	276	MMP-16	348	Prouroguanylin	420	STXBP2	492	Visfatin
61	CD2AP	133	Factor VII	205	ICAM-1	277	MMP-7	349	PRR4	421	SVEP1	493	Vitronectin
62	CD30	134	Factor XII	206	ICAM-2	278	MRP 1	350	PRRC2A	422	SYK	494	WARS
63	CD40 Ligand	135	FAM3C	207	IDE	279	Multimerin 2	351	PRTN3	423	SYN1	495	WISP-1
64	CD9	136	Fas	208	IFN-beta	280	MuSK	352	P-selectin	424	TACI	496	WISP-2
65	CD90	137	Fas Ligand	209	IFNGR1	281	MyBPC3	353	PSMB1	425	TAGLN2	497	XPD
66	CDC14	138	FGF-11	210	IGFBP-2	282	NACA1	354	PSMD2	426	TALDO	498	XPG
67	CFH	139	FGF-20	211	IGSF4C	283	NADK	355	PSMD9	427	TALDO1	499	YY1
68	CFI	140	FGF-23	212	IL-1 alpha	284	NAGPA	356	PSME1	428	Talin-2	500	ZC3H4
69	CFL1	141	FGF-9	213	IL-1 F10	285	NAPRT1	357	PTHLP	429	TARC		
70	CGA	142	FGF-BP	214	IL-1 F5	286	NeuroD1	358	PTMA	430	TARS		
71	CHCHD3	143	FGFR1	215	IL-1 F6	287	Neurolysin	359	PYY	431	TCA-3		
72	Chemerin	144	FGFR2	216	IL-1 F9	288	Neuropilin-2	360	QARS	432	Tcf20		

VIII. Troubleshooting Guide

Problem	Cause	Recommendation
Weak signal	Inadequate detection	Check laser power and PMT parameters
	Inadequate reagent volumes or improper dilution	Check pipettors and ensure correct preparation
	Short incubation times	Ensure sufficient incubation time and change sample incubation step to overnight
	Too low protein concentration in sample	Reduce sample dilution or concentrate sample
	Improper storage of kit	Store kit at suggested temperature
High background	Excess of biotinylated antibodies	Make sure to use the correct amount of antibodies
	Excess of streptavidin	Make sure to use the correct amount of streptavidin
	Inadequate detection	Check laser power and PMT parameters
	Inadequate wash	Increase the volume of wash buffer and incubation time
Uneven signal	Bubbles formed during incubation	Avoid bubble formation during incubation
	Arrays are not completely covered by reagent	Completely cover arrays with solution

IX. Reference List

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